



PD Monitor sample collection kit:

Correct sampling procedures and treatment of blood samples are important to ensure an optimal analysis quality. Please follow our recommendations to ensure you get the best information from your results.

We recommend that the blood sampling is done by experienced staff. To help you get the best from PD Monitor, we have provided a blood sampling kit, which will be replaced after each sample submission. We have also included detailed instructions on taking and handling the necessary samples and packaging for safe transport of the samples to the AFBI laboratory.

Blood sampling (Non-lethal):

Serum from fish blood can be used:

- To detect pre-clinical early SAV (Salmonid Alphavirus) infection, demonstrated by the presence of viraemia (virus in blood);
- As evidence of previous exposure to SAV, demonstrated by the presence of SAV antibody.

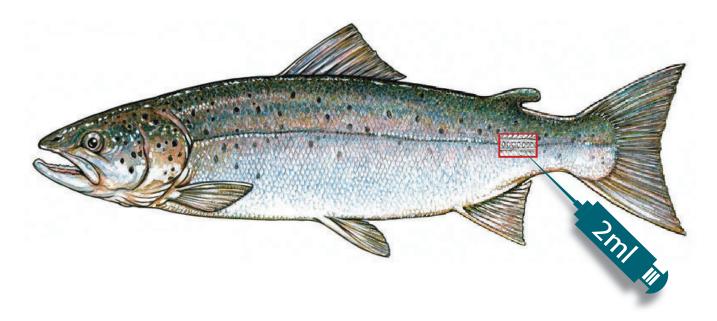
Equipment needed:

- Anaesthetic bath
- Tricaine Pharmag anaesthetic
- 2ml syringe + 21G 1 or 1½ inch needle per fish
- Blood collection and serum transport tubes with labels
- Disposable Pasteur pipettes
- Cool box and needle disposal box

Method:

- Fish should be anaesthetised in small batches in a dilute solution of Tricaine Pharmag.
- An aerated recovery tank should always be available.
- The fish should be placed on its right side or on its back in a V-shaped channel.
- The needle (with the bevel facing the tail) is inserted 5 mm posterior to the anal fin at the mid line at approximately 45° angle to the spine. (See Diagram 1 on page overleaf)
- When the needle touches the spine withdraw it 1mm, blood should pool in the needle hub if the needle is in the caudal vein. Apply light suction with the syringe until enough blood is drawn (1-2ml).
- Remove the needle from the syringe and place whole blood in an Eppendorf tube. (Never discharge blood through a needle as this may cause haemolysis of the blood sample and may make it useless)
- Return the fish to a recovery tank and transfer it to a cage when it is swimming normally.
- Keep all blood samples cool by using cool bags and packs and use one syringe and needle for each fish to prevent cross contamination.
- Always replace the cap on needles and dispose of correctly in a sharps container.
- Draw off the straw coloured serum after centrifugation or after leaving the samples at +4°C overnight using a separate disposable pipette for each sample. Place in clean, labelled Eppendorf tubes.

Location for non-lethal blood sampling from a salmonid fish



Packaging

Place 2 sheets of folded absorbent kitchen paper in the clear plastic zip-lock bag provided before adding the tubes of serum. It is a requirement that absorbent material is included in case of damage or spills during transit. For the same reason, please use the waterproof bubble-lined envelope supplied for the outer wrapping. A coolpak can be added to the package to make sure that the serum samples remain cool in transit.

Paperwork

Please complete fully the information on the address label. This is a legal requirement for sending biological material through the post. Also complete the electronic sample submission form and email to the required recipients (detailed on form). Please also include a hard copy with the samples.

Important

Samples received at AFBI by Thursday each week will be reported the following Monday.

NB An import licence is required for all samples that originate outside Northern Ireland. Please contact us if you require one.

